# STATISTICAL METHODS FOR NEXT GENERATION SEQUENCING

http://www.biostat.jhsph.edu/~khansen/enar2012.html

Zhijin Wu Brown University

Kasper Hansen, Rafael A Irizarry Johns Hopkins University

### OUTLINE

- Introduction to NGS
- SNP calling and genotyping
- RNA-sequencing
- Hands-on exercise

# INTRODUCTION TO NEXT GENERATION SEQUENCING

RAFAEL A. IRIZARRY

http://rafalab.org

Many slides courtesy of: Héctor Corrada Bravo and Ben Langmead

### **REMEMBER THIS?**



D. melanogaster, Science, 2000

H. sapiens, Nature, 2000 and Science, 2000

M. musculus, Nature, 2002

Back then: millions of clones (thousand bps) in 9 months for billions of dollars Today: billion of short reads (35-100 bps) in a week for thousands of dollars Claim: Assemble a genome in weeks for less than \$100,000

## START WITH DNA (MILLIONS OF COPIES)



### **BREAK IT**



### PUT IN SEQUENCER



## SEQUENCE FIRST 35-400 BPS: CALL THEM "READS"

GTTGAGGCTTGCGTTTTTTGGTACGCTGGACTTTGT GTACTCGTCGCTGCGTTGAGGCTTGCGTTTTGGT ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT **TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC** CTTGCGTTTATGGTACGCTGGACTTTGTAGGATAC ͲͲĠĊĠͲͲͲΑͲĠĠͲAĊĠĊͲĠĠAĊͲͲͲĠͲAĠĠAͲAĊĊ GCGTTTATGGTACGCTGGACTTTGTAGGATACCCT GAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGG GCGTTGAGGCTTGCGTTTATGGTACGCTGGATTTT CGTTTATGGTACGCTGGACTTTGTAGGATACCCTC ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT GTTTATGGTACGCTGGACTTTGTAGGATACCCTCG ТСТССТССТССТСССТСССТТСАСССТТСССТТА TGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTA GCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTAC TATGGTACGCTGGACTTTGTAGGATACCCTCGCTT TCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTTTG CGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCT GTTGAGGCTTGCGTTTATGGTACGCTGGGCTTTTT TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC



### ILLUMINA/SOLEXA



- Eight lanes
- ~160M short reads (~50-70 bp) per lane



### NOT JUST ASSEMBLY

- Resequencing
- SNP discovery and genotyping
- Variant discovery and quantification
- TF binding sites: ChIP-Seq
- Gene expression: RNA-Seq
- Measuring methylation

## NOT JUST ASSEMBLY



#### Access

This article is part of Nature's premium content.

Published online 15 October 2008 | Nature 455, 847 (2008) | doi:10.1038/455847a

#### News

The death of microarrays?

High-throughput gene sequencing seems to be stealing a march on microarrays. Heidi Ledford looks at a genome technology facing intense competition.

Heidi Ledford

Faster, cheaper DNA sequencing technology is revolutionizing the burgeoning field of personal genomics. But it is having another, more subtle effect.



### **1000** GENOMES PROJECT



### Genotyping

#### 1000 GENOMES PROJECT DATA RELEASE

#### SNP data downloads and genome browser representing four high coverage individuals

The first set of SNP calls representing the preliminary analysis of four genome sequences are now available to download through the EBI FTP site and the NCBI FTP site. The README file dealing with the FTP structure will help you find the data you are looking for.

The data can also be viewed directly through the 1000 Genomes browser at http://browser.1000genomes.org. Launch the browser and view a sample region here.

More information about the data release can be found in the data section of this web site.

Download the 1000 Genomes Browser Quick Start Guide

Quick start (pdf)

## HUMAN EPIGENOME PROJECT



Methylation

## WHAT TO DO WITH ALL THESE SEQUENCES?

GTTGAGGCTTGCGTTTTTTGGTACGCTGGACTTTGT GTACTCGTCGCTGCGTTGAGGCTTGCGTTTTGGT ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT **TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC** CTTGCGTTTATGGTACGCTGGACTTTGTAGGATAC ͲͲĠĊĠͲͲͲΑͲĠĠͲAĊĠĊͲĠĠAĊͲͲͲĠͲAĠĠAͲAĊĊ GCGTTTATGGTACGCTGGACTTTGTAGGATACCCT GAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGG GCGTTGAGGCTTGCGTTTATGGTACGCTGGATTTT CGTTTATGGTACGCTGGACTTTGTAGGATACCCTC ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT GTTTATGGTACGCTGGACTTTGTAGGATACCCTCG ТСТССТССТССТСССТСССТТСАСССТТСССТТА TGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTA GCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTAC TATGGTACGCTGGACTTTGTAGGATACCCTCGCTT TCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTTTG CGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCT GTTGAGGCTTGCGTTTATGGTACGCTGGGCTTTTT TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC

## MOST APPS: START BY MATCHING TO REFERENCE

GTTGAGGCTTGCGTTTTTGGTACGCTGGACTTTGT

GTACTCGTCGCTGCGTTGAGGCTTGCGTTTTTGGT

ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT

TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC

CTTGCGTTTATGGTACGCTGGACTTTGTAGGATAC

TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC

GCGTTTATGGTACGCTGGACTTTGTAGGATACCCT

GAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGG

GCGTTGAGGCTTGCGTTTATGGTACGCTGGATTTT

CGTTTATGGTACGCTGGACTTTGTAGGATACCCTC

ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT

GTTTATGGTACGCTGGACTTTGTAGGATACCCTCG

TCTCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTA

TGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTA

GCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTAC

TATGGTACGCTGGACTTTGTAGGATACCCTCGCTT

TCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTTTG

CGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCT

GTTGAGGCTTGCGTTTATGGTACGCTGGGCTTTTT

TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC

CTCTCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGGATACCCTCGCTTTC

# Variant detection

#### 

@HWI-EAS146:5:1:1:961#0/1 TCCGAGGCCAACCGAGGCTCCGCGCGCGCTGNN

3>A?B@:@BBBBBAA=BA=A%%%%%%%%%%%%%%%%%%%%% @HWI-EAS146:5:1:1:1595#0/1 TCAGGAAGCAGGAAGAGCTGGTGCAGC

@HWI-EAS146:5:1:1:1048#0/1 CTGGACTGCATCCTACCACCAACTCGTCCAAM

A=B7&7:>B@:A>?9:<;:>?4?%%%%%%%%%%%%%% @HWI-EAS146:5:1:1:1607#0/1 CTCCTCTCAAGGTCCCCAGAAGCACAGCCAA

. BBCCCCCCBBCB7CBC=7>+<>=BCBCB%%%%%%%%%%%%%%% @HWI-EAS146:5:1:1:1719#0/1

CACGATCTGGGTTTATTGTAACCTCCGCCTCNNNNGNTNAAGNNN + BCC?+<B=?BB5=ABA?B6BBBB4BB?B%%%%%%%%%%%%%%%

@HWI-EAS146:5:1:2:947#0/1 CCCAGGAGAAAGCCATGTTCAGTTCGAGCGCNNANANCGTGANNNN

I9@?7A7>AAB@>?B=?@.>8?B?%%%%%%%%%%%%%%%%%%%% @HWI-EAS146:5:1:2:563#0/1 CCAGCCCCTCCCCCATCTCCCACCCTGTACCTNANCCCCTGANNNN

. BBABAABB : AAABA77@5AAA : ??>%%%%%%%%%%%%%%%%%%%%%% @HWI-EAS146:5:1:2:1631#0/1 TGGGAACGCAGCCTACACTCTTCCCAGGCCTCCTNCCTCCGTNN

RRABABBB7:98 @HWI-EAS146:5:1:2:1420#0/1 CTCAAACTCCTGACCTTTGGTGATCCACCCGCCTNGGCCTTCNNNN

BBB:BBBBBABAAA?: (=A8@>AAA?AB?=A%%%%%%%%%%%% @HWI-EAS146:5:1:1:961#0/1 TCCGAGGCCAACCGAGGCTCCGCGGCGCTGN

. BBBB>A?B@:@BBBBBAA=BA=A%%%%%%%%%%%%%%%%%%%%%%%%% @HWI-EAS146:5:1:1:1595#0/1 TCAGGAAGCAGGAAGAGCTGGTGCAGCAG

·RAA20489=1>8888888888888 @HWI-EAS146:5:1:1:1048#0/1 CTGGACTGCATCCTACCACCAACTCGTCCA

. A=B7&7:>B@:A>?9:<::>?4?%%%%%%%%%%%%%%%%%%%%%%% @HWI-EA5146:5:1:1:1607#0/1 CTCCTCTCAAGGTCCCCAGAAGCACAGCCAANNNNANTNNCTNNN

@HWI-EAS146:5:1:1:1719#0/1 CACGATCTGGGTTTATTGTAACCTCCGCCTCNNNNGNTNAAGNN

. BCC?+<B=?BB5=ABA?B6BBBB4BB?B%%%%%%%%%%%%%%% @HWI-EAS146:5:1:2:947#0/1
CCCAGGAGAAAGCCATGTTCAGTTCGAGCGCNNANANCGTGANNNI

@HWI-EAS146:5:1:2:563#0/1 CCAGCCCCTCCCCATCTCCCACCCTGTACCTNANCCCCTGANNNI

@HWI-EAS146:5:1:2:1631#0/1 TGGGAACGCAGCCTACACTCTTCCCAGGCCTCCTNC0





# **RNA-seq differential expression**



# ChIP-seq



### MATCHING REVISTED

GTTGAGGCTTGCGTTTTTGGTACGCTGGACTTTGT

GTACTCGTCGCTGCGTTGAGGCTTGCGTTTTTGGT

ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT

TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC

CTTGCGTTTATGGTACGCTGGACTTTGTAGGATAC

TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC

GCGTTTATGGTACGCTGGACTTTGTAGGATACCCT

GAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGG

GCGTTGAGGCTTGCGTTTATGGTACGCTGGATTTT

CGTTTATGGTACGCTGGACTTTGTAGGATACCCTC

ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT

GTTTATGGTACGCTGGACTTTGTAGGATACCCTCG

TCTCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTA

TGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTA

GCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTAC

TATGGTACGCTGGACTTTGTAGGATACCCTCGCTT

TCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTTTG

CGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCT

GTTGAGGCTTGCGTTTATGGTACGCTGGGCTTTTT

TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC

CTCTCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGGATACCCTCGCTTTC

## MATCHING 10,000,000 32 BPS READS

- BLAST takes more than 6 months
- BLAT takes 2 months
- MAQ takes 1 day and half
- Bowtie takes 17 minutes

### MATCHING

#### GTTGAGGCTTGCGTTTTTGGTACGCTGGACTTTGT GTACTCGTCGCTGCGTTGAGGCTTGCGTTTGCGT

ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC CTTGCGTTTATGGTACGCTGGACTTTGTAGGATAC

#### **Bowtie**

An ultrafast memory-efficient short read aligner

**Bowtie** is an ultrafast, memory-efficient short read aligner. It aligns short DNA sequences (reads) to the human genome at a rate of over 25 million 35-bp reads per hour. Bowtie indexes the genome with a Burrows-Wheeler index to keep its memory footprint small: typically about 2.2 GB for the human genome (2.9 GB for paired-end).



TGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTA GCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTAC TATGGTACGCTGGACTTTGTAGGATACCCTCGCTT TCGTGCTCGCCGCGTTGAGGCTTGCGTTTTTG CGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCT GTTGAGGCTTGCGTTTATGGTACGCTGGGCTTTTT TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC

 ${\tt CTCTCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGGATACCCTCGCTTTC}$ 

# Mapping

#### Take a read:

CTCAAACTCCTGACCTTTGGTGATCCACCCGCCTNGGCCTTC

#### And a reference sequence:

>MT dna:chromosome chromosome:GRCh37:MT:1:16569:1 GATCACAGGTCTATCACCCTATTAACCACTCACGGGAGCTCTCCATGCATTTGGTATTTT CGTCTGGGGGGGTATGCACGCGATAGCATTGCGAGACGCTGGAGCCGGAGCACCCTATGTC GCAGTATCTGTCTTTGATTCCTGCCTCATCCTATTATTATCGCACCTACGTTCAATATT ACAATTGAATGTCTGCACAGCCACTTTCCACACAGACATCATAACAAAAAATTTCCACCA AACCCCCCCTCCCCGCTTCTGGCCACAGCACTTAAACACATCTCTGCCAAACCCCCAAAA ACAAAGAACCCTAACACCAGCCTAACCAGATTTCAAATTTTATCTTTTGGCGGTATGCAC CTCATCAATACAACCCCCGCCCATCCTACCCAGCACA CCCCGAACCAACCAAACCC GCAATACACTGACCCGCTCAAACTCCTGGATTTTGGATC CTAGCCTTTCTATTAGCTCTTAGTA TCACCCTCTAAATCACCACGATCAAAAGGAACAAGCATCAAGCACGCAGCAATGCAGCTC AAAACGCTTAGCCTAGCCACACCCCCACGGGAAACAGCAGTGATTAACCTTTAGCAATAA GGTCACACGATTAACCCAAGTCAATAGAAGCCGGCGTAAAGAGTGTTTTAGATCACCCCC TCCCCAATAAAGCTAAAACTCACCTGAGTTGTAAAAAACTCCAGTTGACACAAAATAGAC TACGAAAGTGGCTTTAACATATCTGAACACACAATAGCTAAGACCCCAAACTGGGATTAGA TACCCCACTATGCTTAGCCCTAAACCTCAACAGTTAAATCAACAAAACTGCTCGCCAGAA CACTACGAGCCACAGCTTAAAACTCAAAGGACCTGGCGGTGCTTCATATCCCTCTAGAGG AGCCTGTTCTGTAATCGATAAACCCCGATCAACCTCACCACCTCTTGCTCAGCCTA CCGCCATCTTCAGCAAACCCTGATGAAGGCTACAAAGTAAGCGCAAGTACCCACG ACGTTAGGTCAAGGTGTAGCCCATGAGGTGGCAAGAAATGGGCTACATTTTCTAC AAAACTACGATAGCCCTTATGAAACTTAAGGGTCGAAGGTGGATTTAGCAGTAMACTAAG AGTAGAGTGCTTAGTTGAACAGGGCCCTGAAGCGCGTACACACCGCCCGTCACCCTCCTC AAGTATAC CGTAACTCCAAACTCCTGCCTTTGGTGATCCACCCGCCTTGGCCTACCTGCATAATGAAG GCCCCAAACCCACTCCACCTTACTACCAGACAACCTTAGCCAAACCATTTACCCAAATAA AGTATAGGCGATAGAAATTGAAACCTGGCGCAATAGATATAGTACCGCAAGGGAAAGATG AAAAATTATAACCAAGCATAATATAGCAAGGACTAACCCCTATACCTTCTGCATAATGAA TTAACTAGAAATAACTTTGCAAGGAGAGCCCAAAGCTAAGACCCCCGAAACCAGACGAGCT

How do we determine the read's point of origin with respect to the reference?

Answer: sequence similarity



Say hypothesis 2 is correct. Why are there still mismatches and gaps?

### More on variants and base-calling

### **SNPs**

GTTGAGGCTTGCGTTTTTTGGTACGCTGGACTTTGT

**GTACTCGTCGCTGCGTTGAGGCTTGCGTTTTTGGT** 

**A**TGGTACGCTGGACTTTGTAGGATACCCTCGCTTT

TTGCGTTT<mark>A</mark>TGGTACGCTGGACTTTGTAGGATACC

CTTGCGTTT<mark>A</mark>TGGTACGCTGGACTTTGTAGGATAC

TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC

GCGTTTATGGTACGCTGGACTTTGTAGGATACCCT

GAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGG

GCGTTGAGGCTTGCGTTTATGGTACGCTGGATTTT

CGTTTATGGTACGCTGGACTTTGTAGGATACCCTC

**ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT** 

GTTTATGGTACGCTGGACTTTGTAGGATACCCTCG

TCTCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTA

TGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTA

GCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTAC

TATGGTACGCTGGACTTTGTAGGATACCCTCGCTT

TCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTTTG

CGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCT

**GTTGAGGCTTGCGTTTATGGTACGCTGGGCTTTTT** 

TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC

CTCTCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGGATACCCTCGCTTTC

### **SNPs**

TCTCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTA TCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTTTG **GTACTCGTCGCTGCGTTGAGGCTTGCGTTTTTGGT** ͲĠĊͲĊĠͲĊĠĊͲĠĊĠͲͲĠĂĠĠĊͲͲĠĊĠͲͲͲĂͲĠĠͲĂ GCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTAC CGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCT GCGTTGAGGCTTGCGTTTATGGTACGCTGGATTTT GTTGAGGCTTGCGTTTTTGGTACGCTGGACTTTGT GTTGAGGCTTGCGTTTATGGTACGCTGGGCTTTTT GAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGG CTTGCGTTTATGGTACGCTGGACTTTGTAGGATAC TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC TTGCGTTTATGGTACGCTGGACTTTGTAGGATACC GCGTTTATGGTACGCTGGACTTTGTAGGATACCCT CGTTTATGGTACGCTGGACTTTGTAGGATACCCTC GTTTATGGTACGCTGGACTTTGTAGGATACCCTCG TATGGTACGCTGGACTTTGTAGGATACCCTCGCTT **ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT ATGGTACGCTGGACTTTGTAGGATACCCTCGCTTT** 

CTCTCGTGCTCGTCGCTGCGTTGAGGCTTGCGTTTATGGTACGCTGGACTTTGTAGGATACCCTCGCTTTC



### 1000 Genomes Data

#### SNPs in dbSNP



### 1000 Genomes Data

SNPs in dbSNP

#### Novel SNPs



# What is causing this?



### Before Reads There were Intensities

<pre>&gt; ints[1:10,1:4]</pre>				
	A.1	C.1	G.1	T.1
1	154.8	122.1	119.3	13001.9
2	1093.5	6186.6	-798.4	208.3
3	892.3	4028.2	-367.9	-463.9
4	590.5	2607.9	-81.6	188.7
5	979.4	6411.0	943.5	454.9
6	945.5	4943.1	19.7	-1170.8
7	255.0	213.3	15.5	4358.8
8	1085.2	5834.5	-384.7	-94.1
9	267.6	340.3	6866.2	5788.6
10	1162.6	6424.4	-497.6	-149.2

### We Want to See This

![](_page_33_Figure_1.jpeg)

Four-channel fluorescence intensity, cycle 1

Color coded by call made: A, C, G, T

![](_page_34_Figure_0.jpeg)

### Gets Worse for higher cycles

![](_page_35_Figure_1.jpeg)

### Error Rate and Reported Quality

![](_page_36_Figure_1.jpeg)

### Remember This?

![](_page_37_Figure_1.jpeg)

![](_page_38_Figure_0.jpeg)

### Base Calling

1) Rougemont et al. Probabilistic base calling of Solexa sequencing data. BMC Bioinformatics (2008)

2) Erlich et al. Alta-Cyclic: a self-optimizing base caller for next-generation sequencing. Nat Methods (2008)

3) Kao et al. BayesCall: A model-based base-calling algorithm for high-throughput short-read sequencing. Genome Res (2009)

4) Corrada Bravo and Irizarry. Model-Based Quality Assessment and Base-Calling for Second-Generation Sequencing Data. Biometrics (2009)

5) Cokus et al. Shotgun bisulphite sequencing of the Arabidopsis genome reveals DNA methylation patterning. Nature (2009)

![](_page_40_Figure_1.jpeg)

cycle

#### log intensity read i, cycle j, channel c

$$u_{ijc} = \underline{\Delta_{ijc}}(\mu_{cj\alpha} + x_j^T \alpha_i + \epsilon_{ijc}^{\alpha}) + \underline{(1 - \Delta_{ijc})}(\mu_{cj\beta} + x_j^T \beta_i + \epsilon_{ijc}^{\beta})$$

### indicators of nucleotide identity, read i, pos. j

$$\Delta_{ijc} = \begin{cases} 1 & \text{if } c \text{ is the nucleotide in read } i \text{ position } j \\ 0 & \text{otherwise} \end{cases}$$

#### log intensity read *i*, cycle *j*, channel *c*

$$u_{ijc} = \Delta_{ijc}(\mu_{cj\alpha} + \underline{x_j^T \alpha_i} + \epsilon_{ijc}^{\alpha}) + (1 - \Delta_{ijc})(\mu_{cj\beta} + \underline{x_j^T \beta_i} + \epsilon_{ijc}^{\beta})$$

read-specific linear models

#### log intensity read *i*, cycle *j*, channel *c*

$$u_{ijc} = \Delta_{ijc}(\mu_{cj\alpha} + x_j^T \alpha_i + \underline{\epsilon}_{ijc}^{\alpha}) + (1 - \Delta_{ijc})(\mu_{cj\beta} + x_j^T \beta_i + \underline{\epsilon}_{ijc}^{\beta})$$

measurement error

 $\epsilon^{\alpha}_{ijc} \sim N(0, \sigma^2_{\alpha i})$ 

 $\epsilon_{ijc}^{\beta} \sim N(0, \sigma_{\beta i}^2)$ 

### Read & Cycle Effects

![](_page_44_Figure_1.jpeg)

cycle

![](_page_45_Figure_0.jpeg)

![](_page_46_Figure_0.jpeg)

### The End