





- Each gene or portion of a gene is represented by 11 to 20 oligonucleotides of 25 base-pairs.

- Reporter/Feautre/Probe: an oligonucleotide of 25 base-pairs, i.e., a 25-mer. Perfect match (PM): A 25-mer complementary to a reference sequence of interest (e.g., part of a gene). Mismatch (MM): same as PM but with a single homomeric base change for the middle (13^m) base (transversion purine <> pyrimidine, G <> C, A <> T). Probe-pair: a (PM,MM) pair. Probe-pair set: a collection of probe-pairs (11 to 20) related to a common gene or fraction of a gene. Affy ID: an identifier for a probe-pair set. The purpose of the MM probe design is to measure non-specific binding and background noise.

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Affymetrix files

- Main software from Affymetrix company MicroArray Suite - MAS, now version 5.
- DAT file: Image file, ~10^7 pixels, ~50 MB.
- CEL file: Cell intensity file, probe level PM and MM values.
- CDF file: Chip Description File. Describes which probes go in which probe sets and the location of probe-pair sets (genes, gene fragments, ESTs).

Expression Measures

- 10-20K genes represented by 11-20 pairs of probe intensities (PM & MM)
- Obtain expression measure for each gene on each array by summarizing these pairs
- We already discussed background adjustment and normalization. We assume this has been done.
- · There are many methods

Data and notation

- *PM_{ijg}*, *MM_{ijg}* = Intensity for perfect match and mismeth probe in cell *j* for gene *g* in chip *i*.
 - i = 1,..., n -- from one to hundreds of chips; - j = 1,..., J -- usually 11 or 20 probe pairs;
 - -g = 1,..., G -- between 8,000 and 20,000 probe sets.
- Task: summarize for each probe set the probe level data, i.e., PM and MM pairs, into a single expression measure.
- Expression measures may then be compared within or between chips for detecting differential expression.

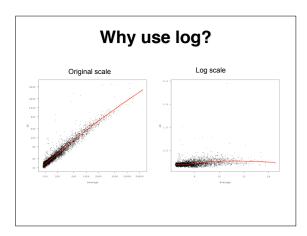
MAS 4.0

 GeneChip[®] MAS 4.0 software used AvDiff up until 2001

$$AvDiff = \frac{1}{|A|} \sum_{j \in A} (PM_j - MM_j)$$

where A is a set of "suitable" pairs, e.g., pairs with $d_j = PM_j - MM_j$ within 3 SDs of the average of $d_{(2)}, ..., d_{(J-1)}$

- Obvious problems:
 - Negative values
 - No log scale

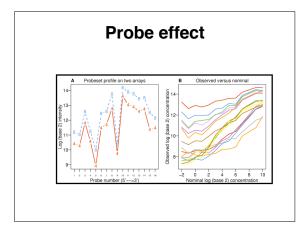




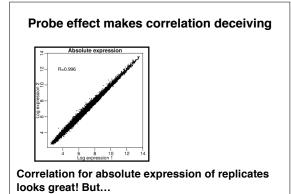
Li and Wong's observations

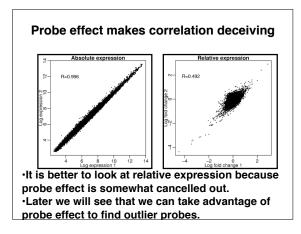
- There is a large probe effect
- There are outliers that are only noticed when looking across arrays
- Non- linear normalization needed (discussed in previous lecture)

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Li & Wong

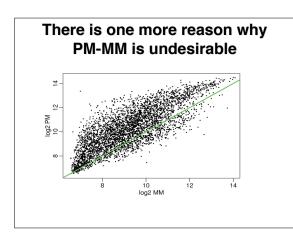
• Li & Wong (2001) fit a model for each probe set, i.e., gene

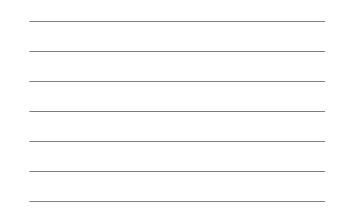
 $PM_{ij} - MM_{ij} = \theta_i \phi_j + \varepsilon_{ij}, \ \varepsilon_{ij} \propto N(0,\sigma^2)$

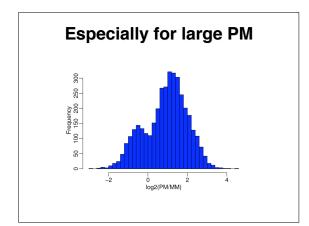
where

- θ_i: model based expression index (MBEI),
 φ_i: probe sensitivity index.
 Maximun likelihood estimate of MBEI is used as expression measure for the gene in chip *i*.
 Non-linear normalization used

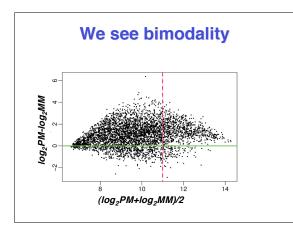
- Ad-hoc procedure used to remove outliers
- Need at least 10 or 20 chips



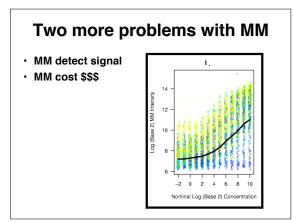










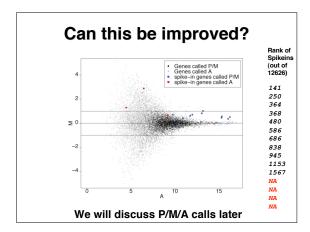


MAS 5.0

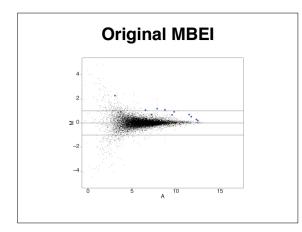
Current version, MAS 5.0, uses Signal

 $signal = Tukey Biweight\{log(PM_j - MM_j^*)\}$

- Notice now log is used
- · But what about negative PM-MM ?
- MM ^x is a new version of MM that is never larger than PM.
 If MM < PM, MM⁺ = MM.
 If MM >= PM,
 SB = Tukey Biweight (log(PM)-log(MM))
 (log-ratio).
 - log(MM)+ = log(PM)-log(max(SB, +ve)).
 Tukey Biweight: B(x) = (1 (x/c)^2)^2 if IxIcc, 0 ow.





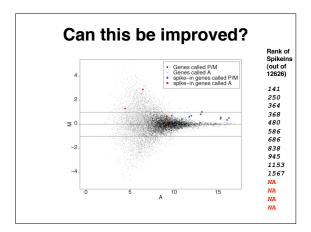


RMA

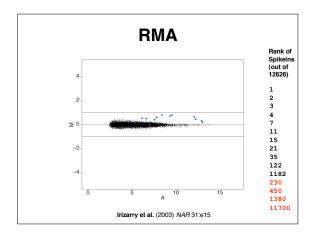
- Robust regression method to estimate expression measure and SE from PM' (background adjusted normalized PM)
- Use quantile normalization
- Assume additive model

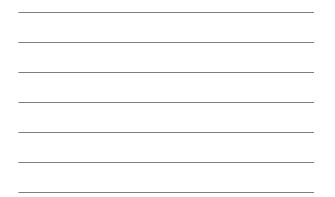
$\log_2(PM^*_{ij}) = a_i + b_j + \varepsilon_{ij}$

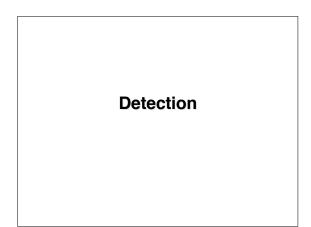
- Estimate RMA = a_i for chip i using robust method, such as median polish (fit iteratively, successively removing row and column medians, and accumulating the terms, until the process stabilizes).
- Works with *n=2* or more chips
 This is a robust multi-array analysis (RMA)











Detection

- The detection problem: "Given the probe-level data, which mRNA transcripts are present in the sample?"
- Biologists are mostly interested in expression levels, and so detection has received less attention
- $\boldsymbol{\cdot}$ To date only Affymetrix has tackled this, with
 - Rank-based tests
 - Implemented in MAS5.0

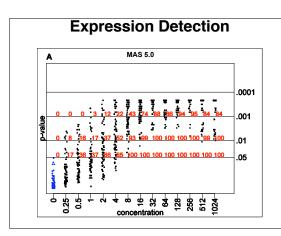
MAS Rank-based Detection

The test used in MAS 5.0 compares the following two hypotheses

$$\begin{split} H_0\colon median\;(PM_j\cdot MM_j)/(PM_j+MM_j) &= \tau\;;\\ H_1\colon median\;(PM_j\cdot MM_j)/(PM_j+MM_j) > \tau. \end{split}$$

Significance levels: $0 < \alpha_1 < \alpha_2 < 0.5$. If p is the p-value for the (rank) test, MAS 5.0 calls a transcript absent: if $p > \alpha_2$, marginal: if $\alpha_1 \leq p \leq \alpha_2$, and present: if $p < \alpha_1$.

Typically tests are carried out with $\,\tau$ = 0.15, $\,\alpha_1$ = .04 and α_2 = .06.



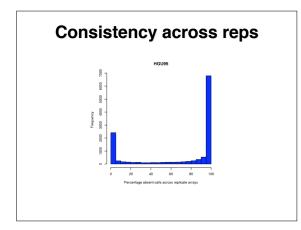


Remember uncertainty

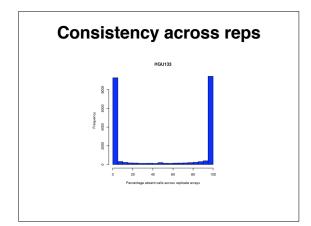
- Some data analysts remove probesets called absent from further analysis
- This creates false negatives:

P	M	Α
82%	1%	17%
0%	0%	100%
Р	М	Α
77%	3%	20%
0%	0%	100%
	0% P 77%	O% O% P M 77% 3% 0% 0%











Current work

- We need better estimates of means and variances of bivariate normal background noise
- Use observed MM intensities along with sequence information
- We also have a solution that does not use the MM